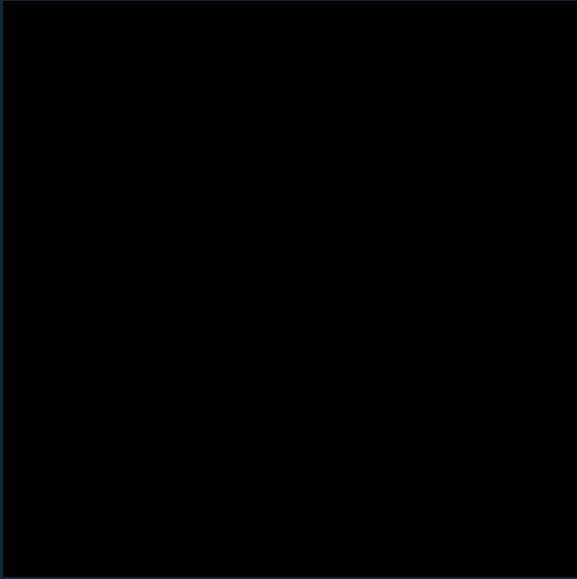
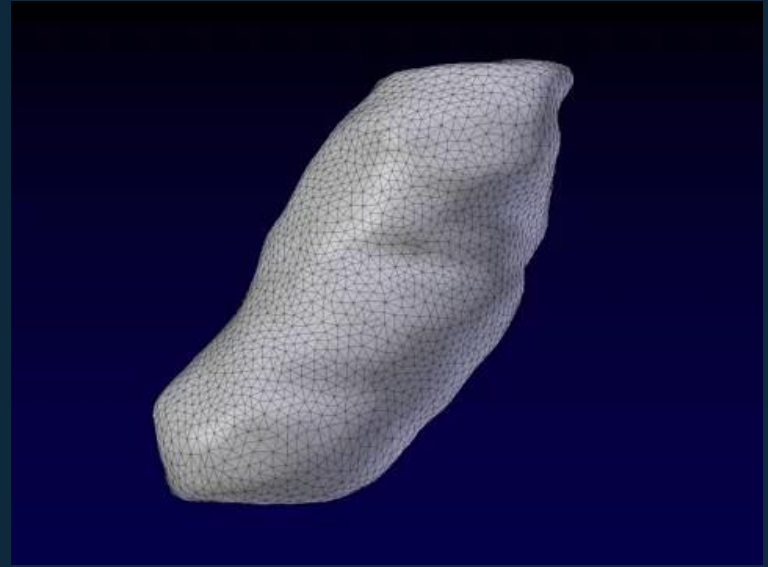


Design principles of mitochondrial architecture in neurons

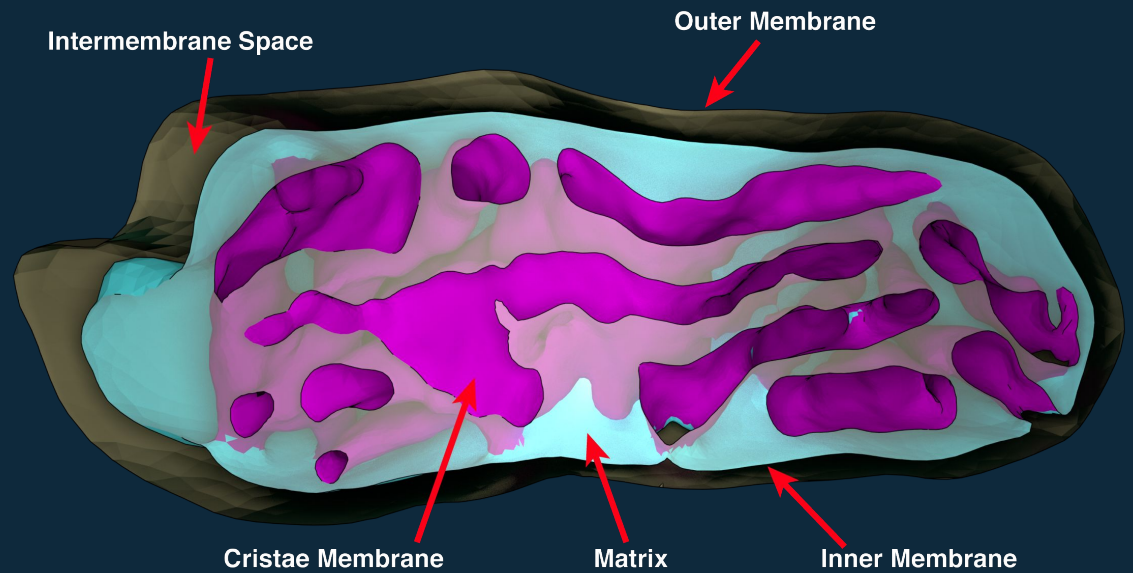


Rachel Mendelsohn
Sejnowski Lab
Salk Institute



Christopher T. Lee
Rangamani and Holst Labs
University of California San Diego

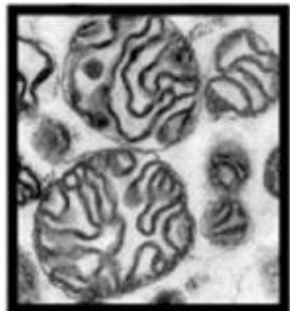
Dual membranes are key for mitochondrial function



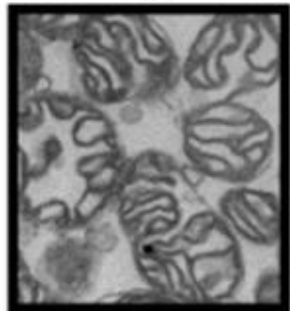
Four Steps for ATP:



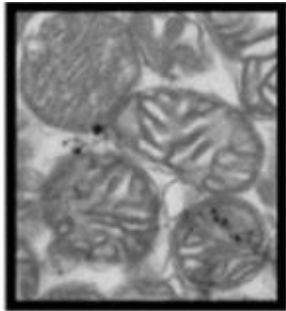
cerebrum



cerebellum



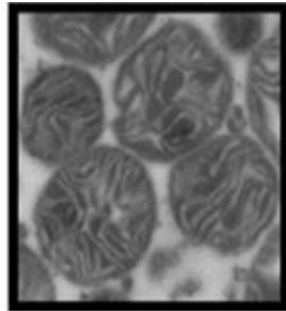
brain stem



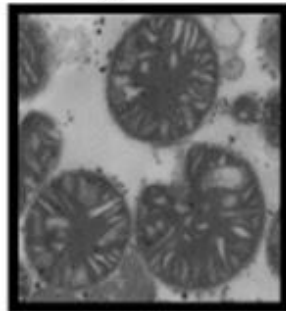
spinal cord



kidney



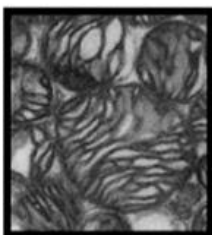
liver



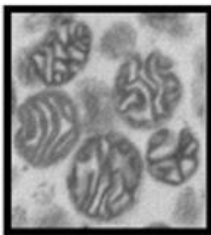
heart



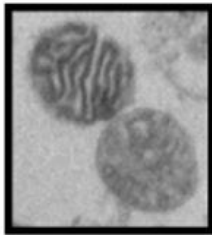
sk muscle



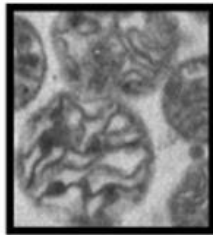
adipose



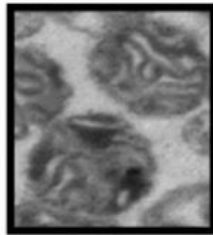
sm intestine



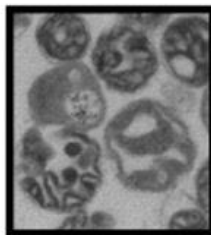
lg Intestine



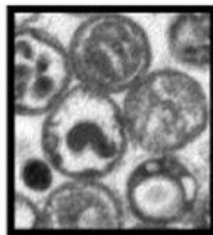
stomach



placenta



testes



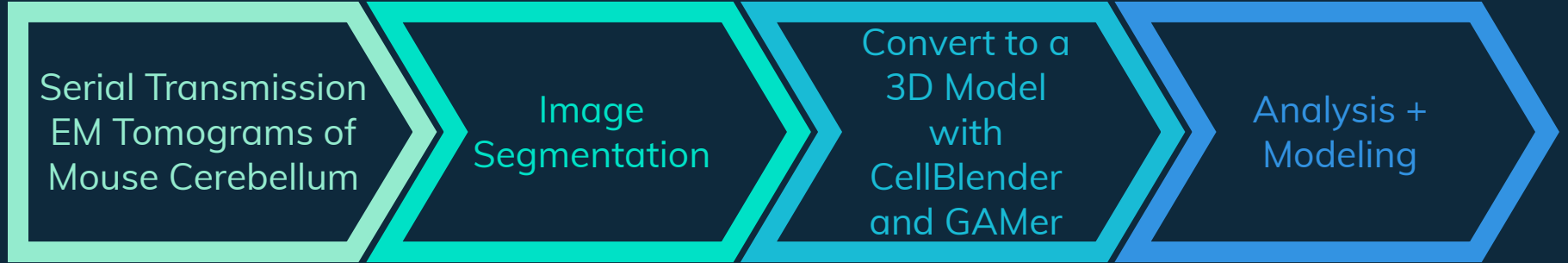


Specificity Suggests Specialization

- ◇ Unique mitochondrial structures reflect unique bioenergetic needs.
- ◇ Neurons have, arguably, the most specialized mitochondria of any cell type.
- ◇ Information about neuronal energy dynamics is contained in mitochondrial structure.



Enabling morphological and modeling analysis with real geometries constrained by EM tomography



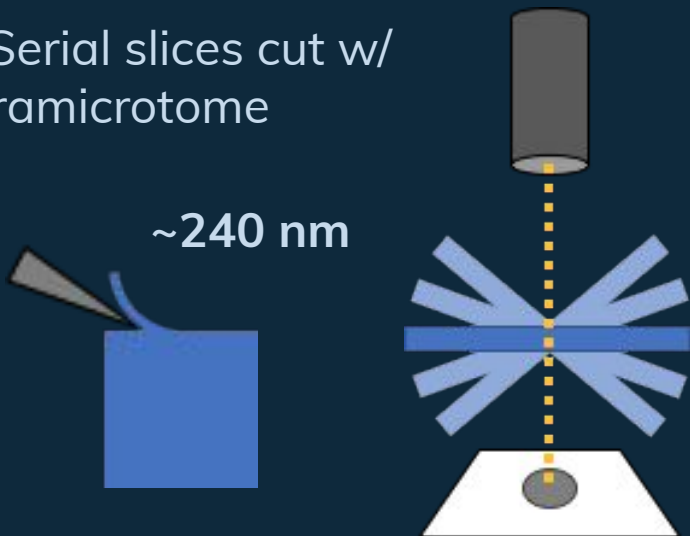
Serial Transmission
EM Tomograms of
Mouse Cerebellum

Image
Segmentation

Convert to a
3D Model
with
CellBlender
and GAMer

Analysis +
Modeling

1. Serial slices cut w/
ultramicrotome



2x Datasets:

$\sim 2.4 \times 2.5 \times 1 \text{ } \mu\text{m}^3$

$\sim 2.4 \times 2.4 \times 1.6 \text{ } \mu\text{m}^3$

1.64 nm isotropic virtual voxel size

~ 35 Total Mitochondria

18 Full

17 Partial

Eric Bushong, Sebastian Phan, Ellisman Lab

2. Slices imaged using multi-tilt
electron tomography



Serial Transmission
EM Tomograms of
Mouse Cerebellum

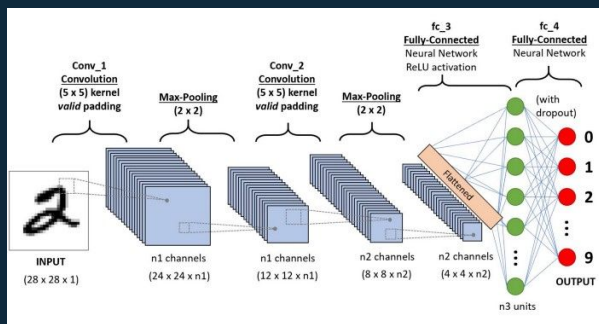
Image
Segmentation

Convert to a
3D Model
with
CellBlender
and GAMer

Analysis +
Modeling

Semi-automated Machine Learning:

- ◇ iLastik (<https://ilastik.org>)
- ◇ CDeep3m (NCMIR)
- ◇ Other tools

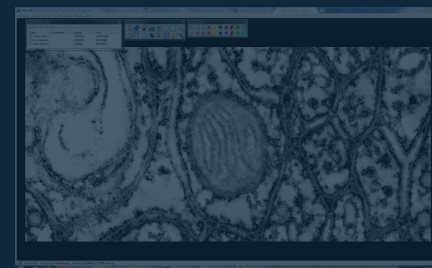


<https://towardsdatascience.com/>

Manual Tracing:

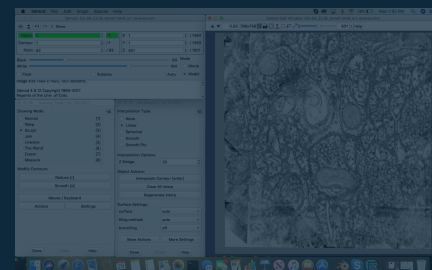
Reconstruct

- ◇ Developed by Kristen Harris Lab (UT Austin)



IMOD

- ◇ Developed by Boulder laboratory for 3-D EM of cells



Object Classification Using Random Forest NN in iLastik

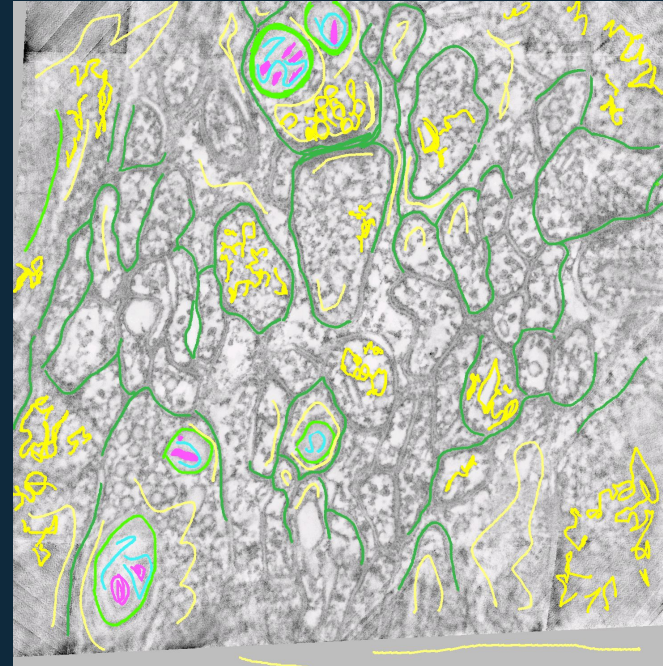
Step 1:

Manually label components to
train Random Forest Classifier

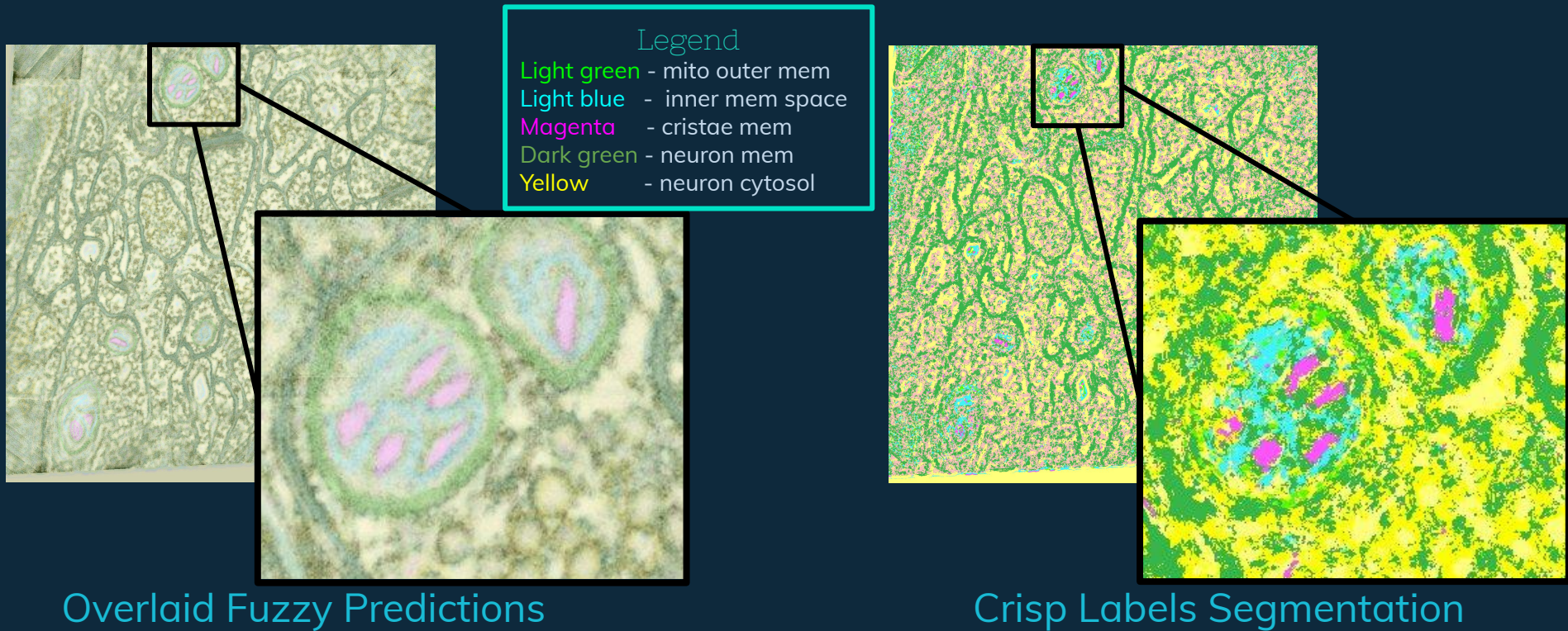


Legend

- Light green - mito outer mem
- Light blue - inner mem space
- Magenta - cristae mem
- Dark green - neuron mem
- Yellow - neuron cytosol



Segmentations from iLastik are unsuitable for mesh generation



Serial Transmission
EM Tomograms of
Mouse Cerebellum

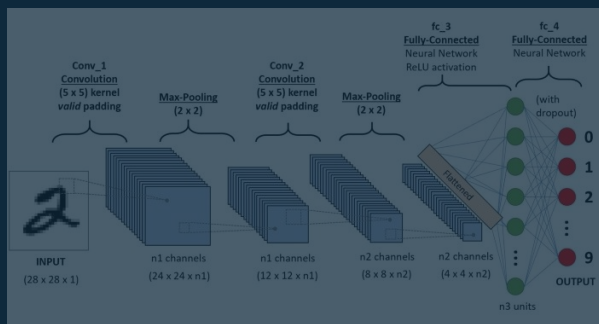
Image
Segmentation

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Modeling

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- ◇ iLastik (<https://ilastik.org>)
- ◇ CDeep3m (NCMIR)
- ◇ Other tools



<https://towardsdatascience.com/>

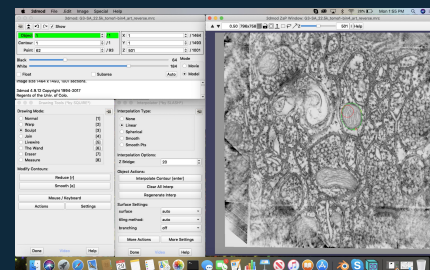
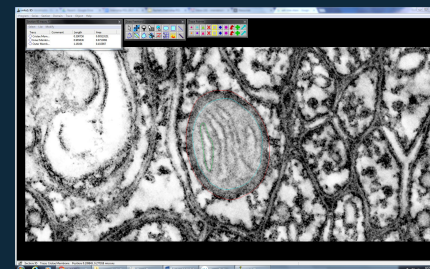
Manual Tracing:

Reconstruct

- ◇ Developed by Kristen Harris Lab (UT Austin)
- ◇ Designed for image segmentation

IMOD

- ◇ Developed by Boulder laboratory for 3-D EM of cells
- ◇ Cross platform modeling suite focused on EM reconstructions



Serial Transmission
EM Tomograms of
Mouse Cerebellum

Image
Segmentation

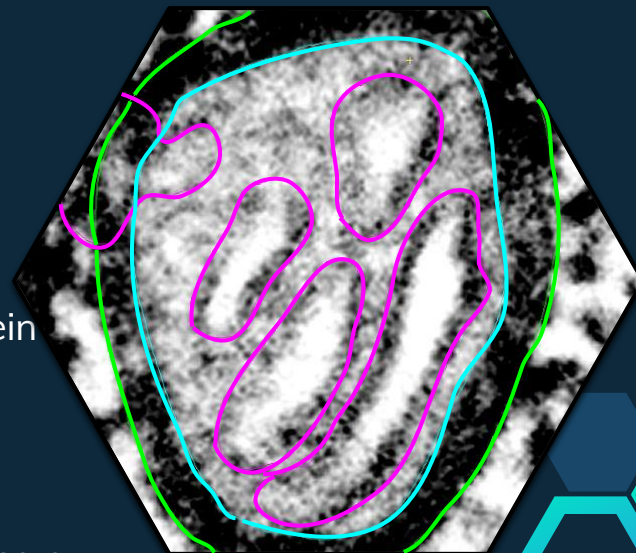
Convert to a
3D Model
with
CellBlender
and GAMer

Analysis +
Modeling

IMOD and Reconstruct
Manual tracing

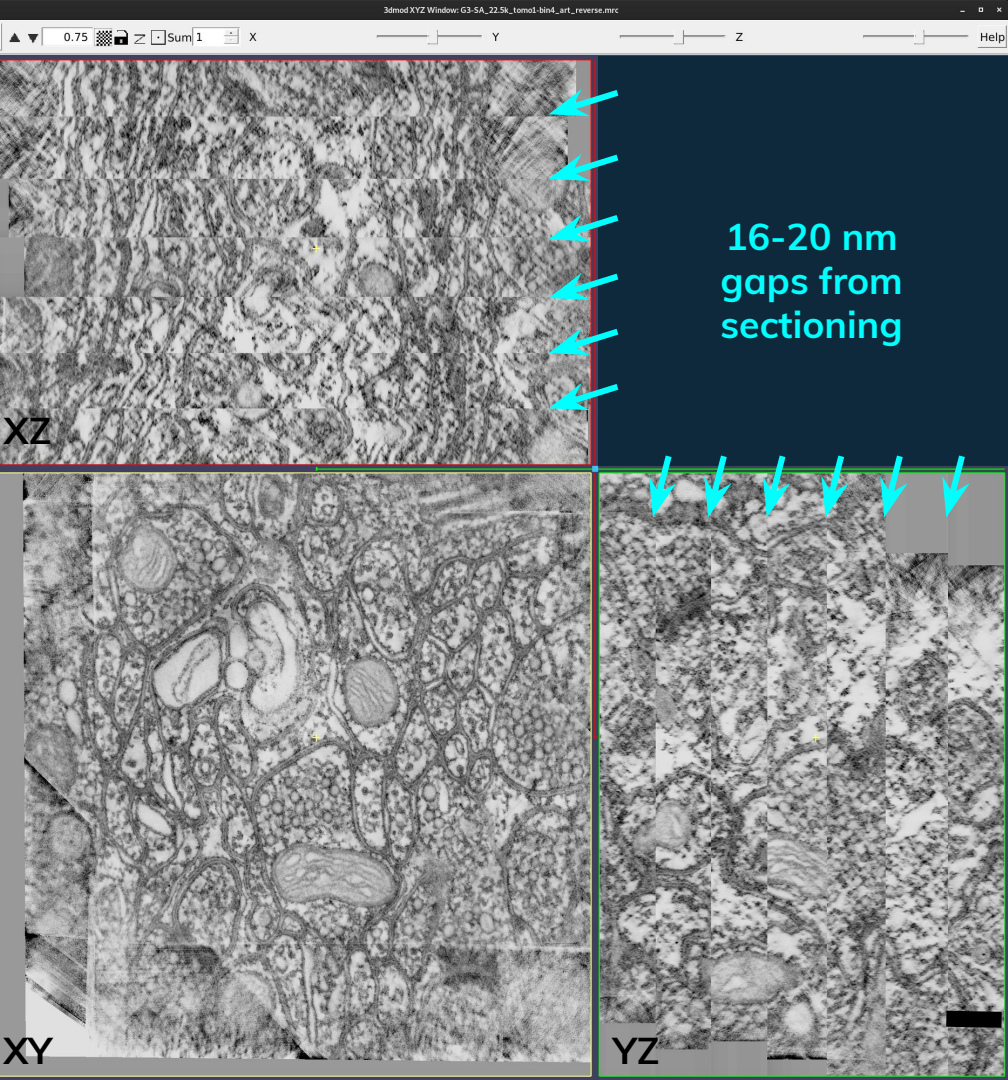
General Guidelines for Tracing:

- 1) Trace **Outer**, **Inner**, **Cristae** separately.
- 2) Create a smooth outer membrane, excluding potential protein structures on the surface
- 3) Trace the matrix side of the inner and cristae membrane (Boundary later moved in Blender)
- 4) Include cristae junction curvature in the cristae trace, creating a smooth intersection with the inner membrane trace



Mito 4





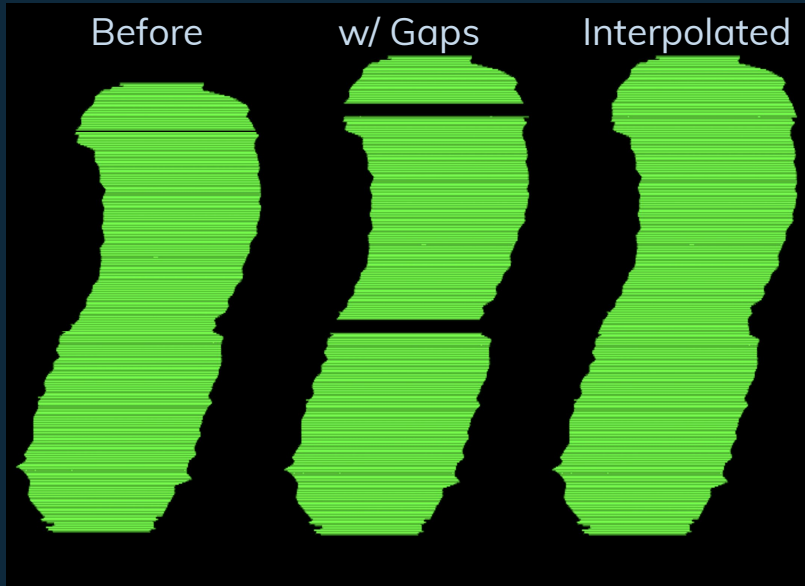
Missing data occurs between slices

- ◇ Knife Removes Material Between Tomograms
- ◇ We estimate 16-20 nm of material lost
- ◇ Contours across gaps can be manually interpolated in IMOD

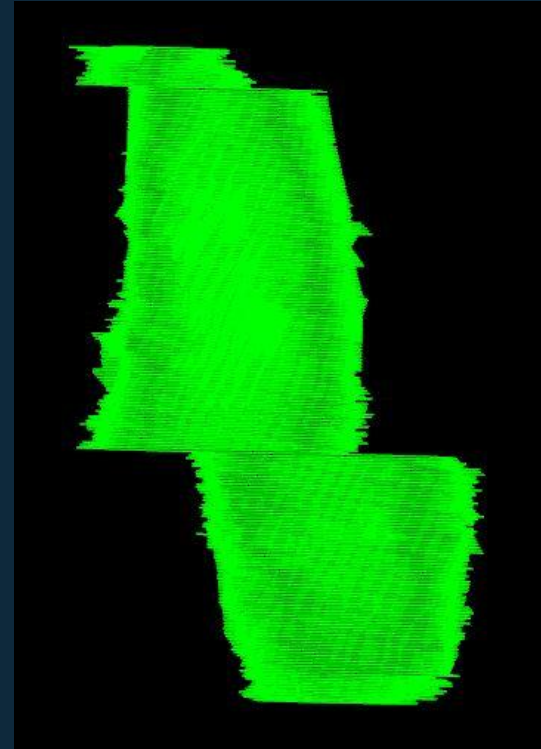
How did we do gap interpolation?

Our Approach...

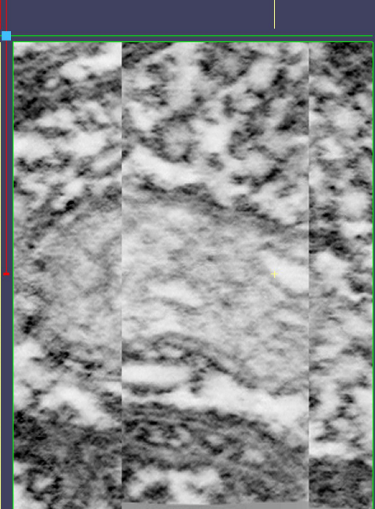
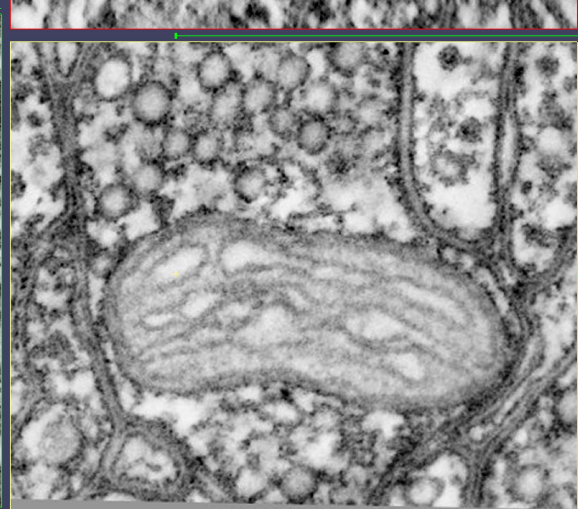
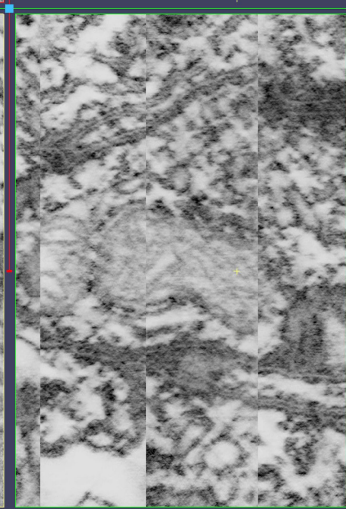
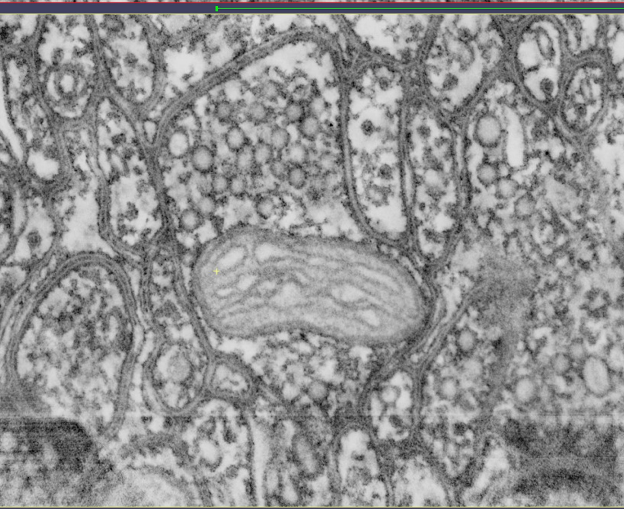
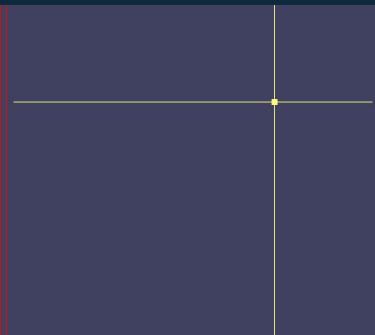
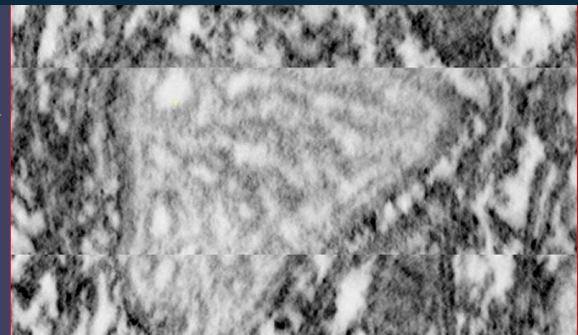
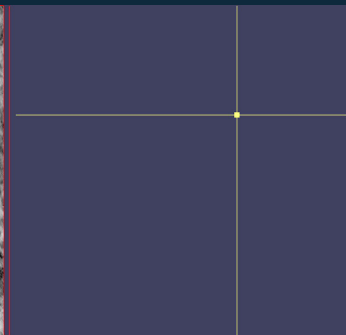
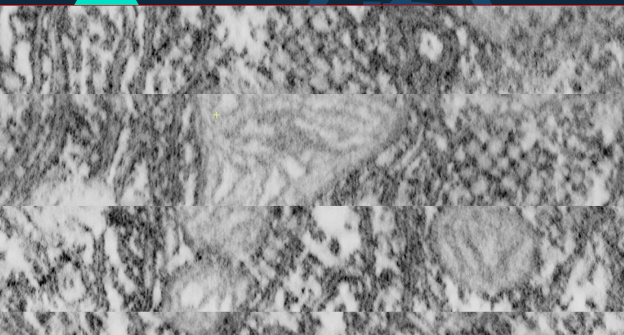
- ◇ Must be Manually Interpolated with the Help of IMOD's Interpolator



Mito 4



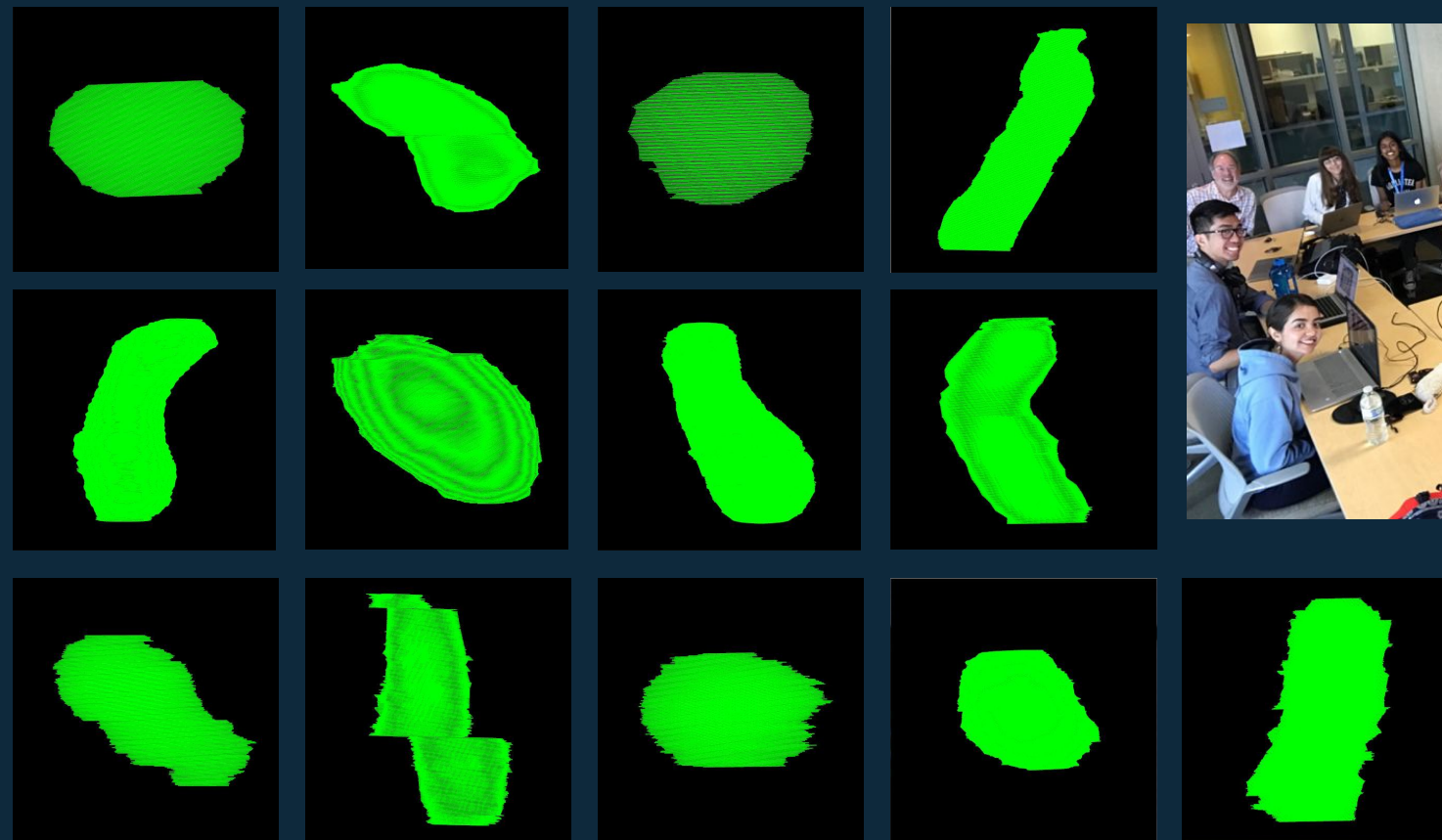
Mito 7



Alignment



Summer 2019 Mitochondria Meshathon

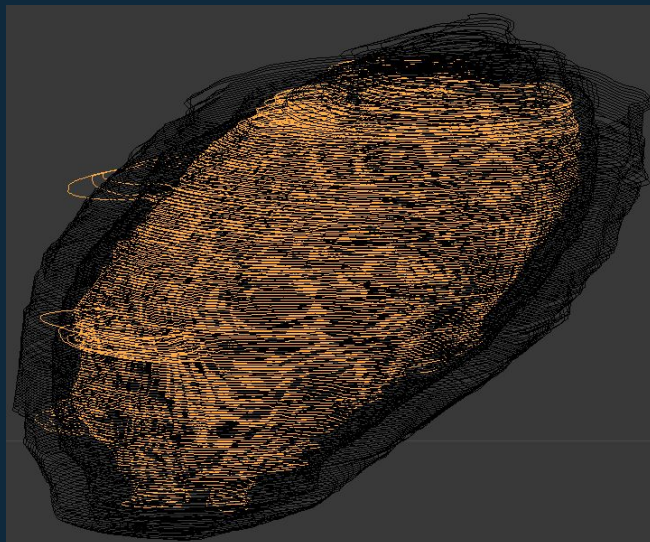
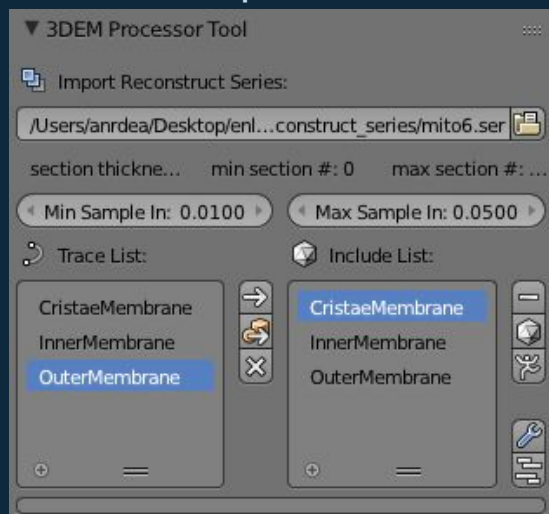


+3

Mitochondria
Segmented in
Reconstruct

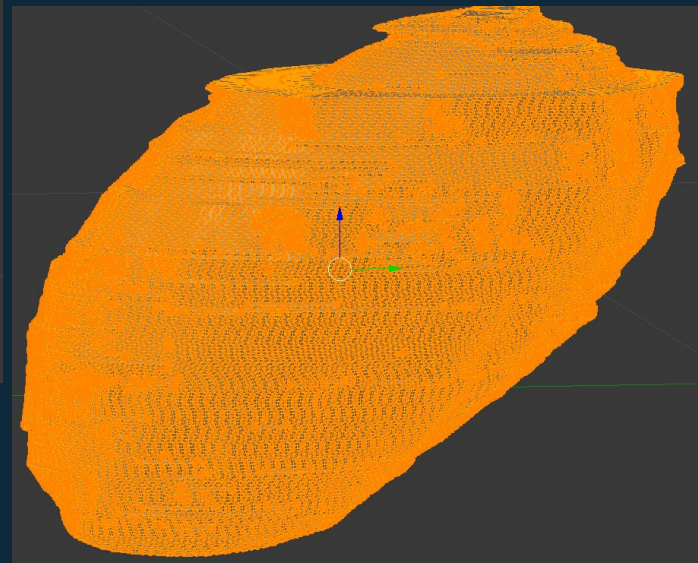
Contour Tiling IMOD/Reconstruct to Blender Mesh

Neuropil Tools

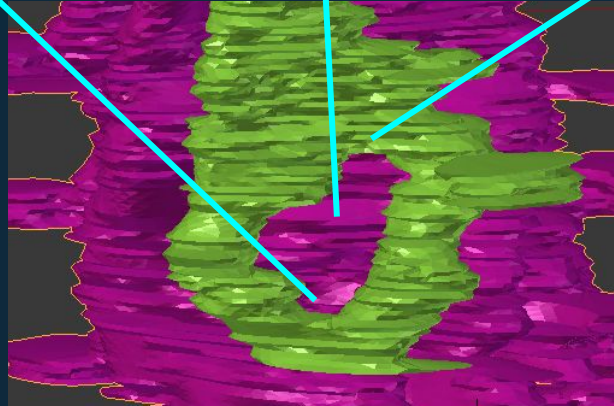
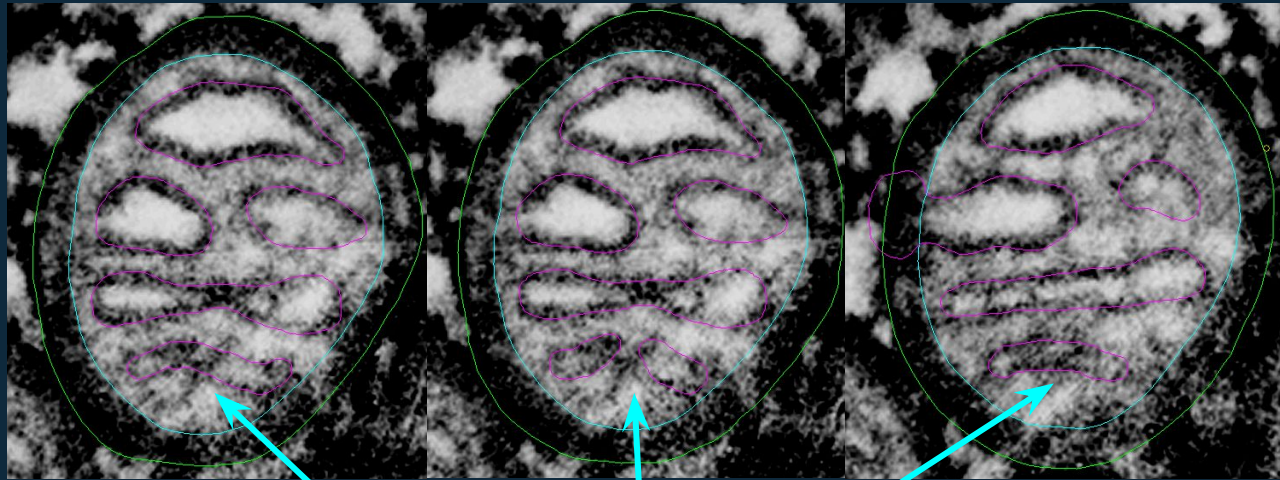


Contours

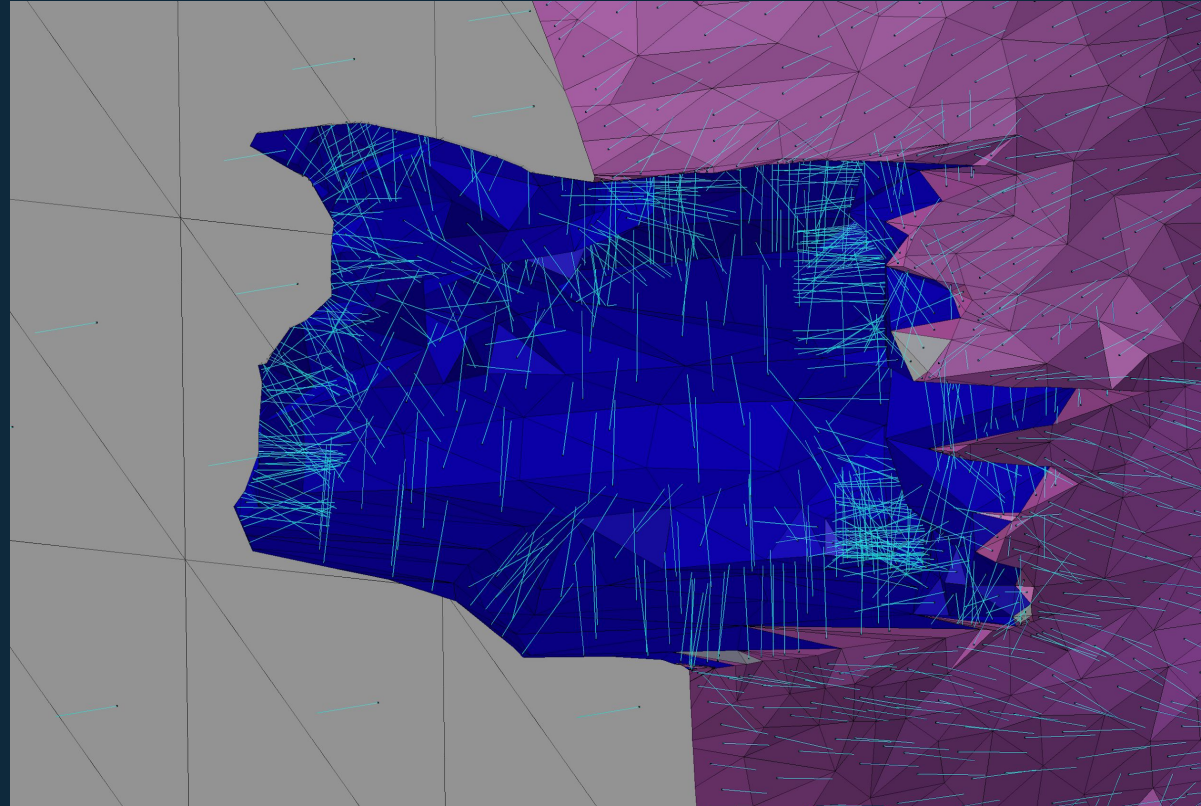
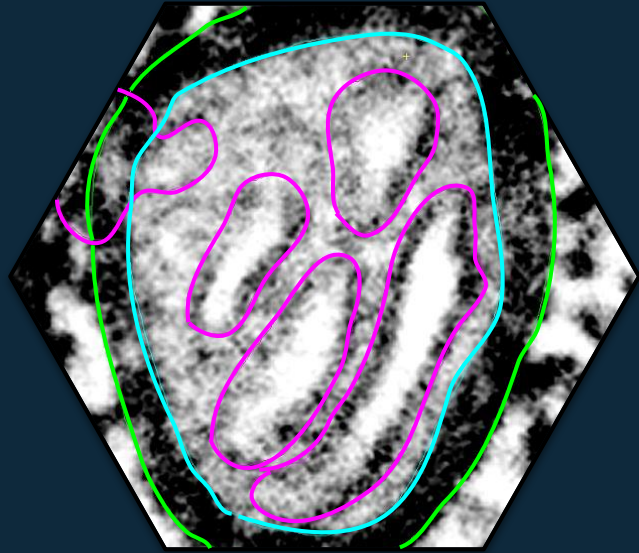
Tiled Mesh of
Outer Membrane



Disappearing EM contrast leads to ambiguity



Anisotropic smoothness of cristae junctions XY vs Z

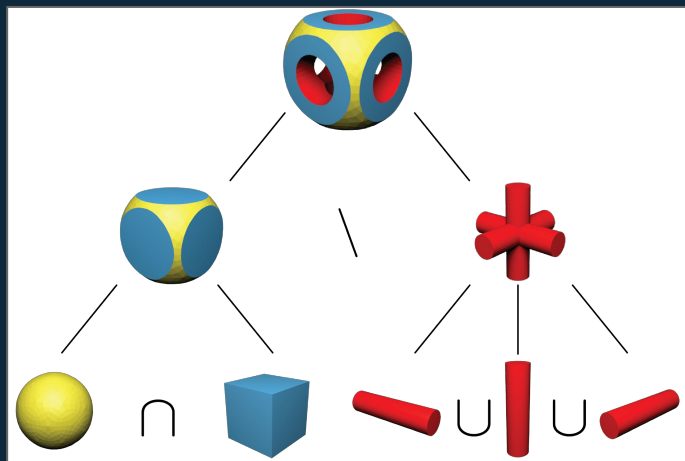


Serial Transmission
EM Tomograms of
Rat Cerebellum

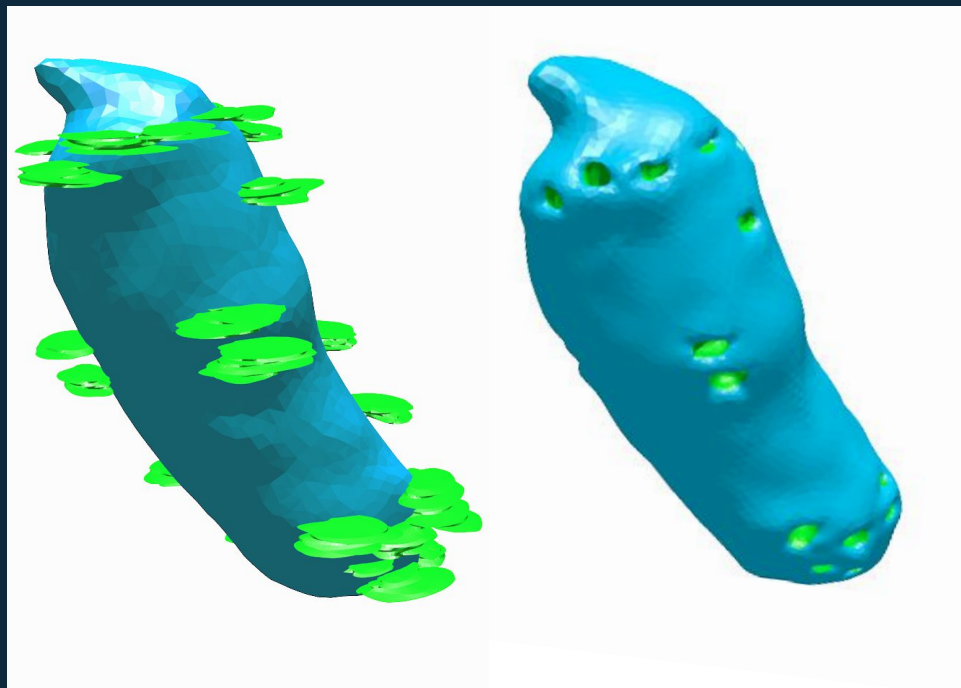
Image
Segmentation

Convert to a
3D Model
with
CellBlender
and GAMer

Analysis +
Modeling



Geometric Boolean
Operations

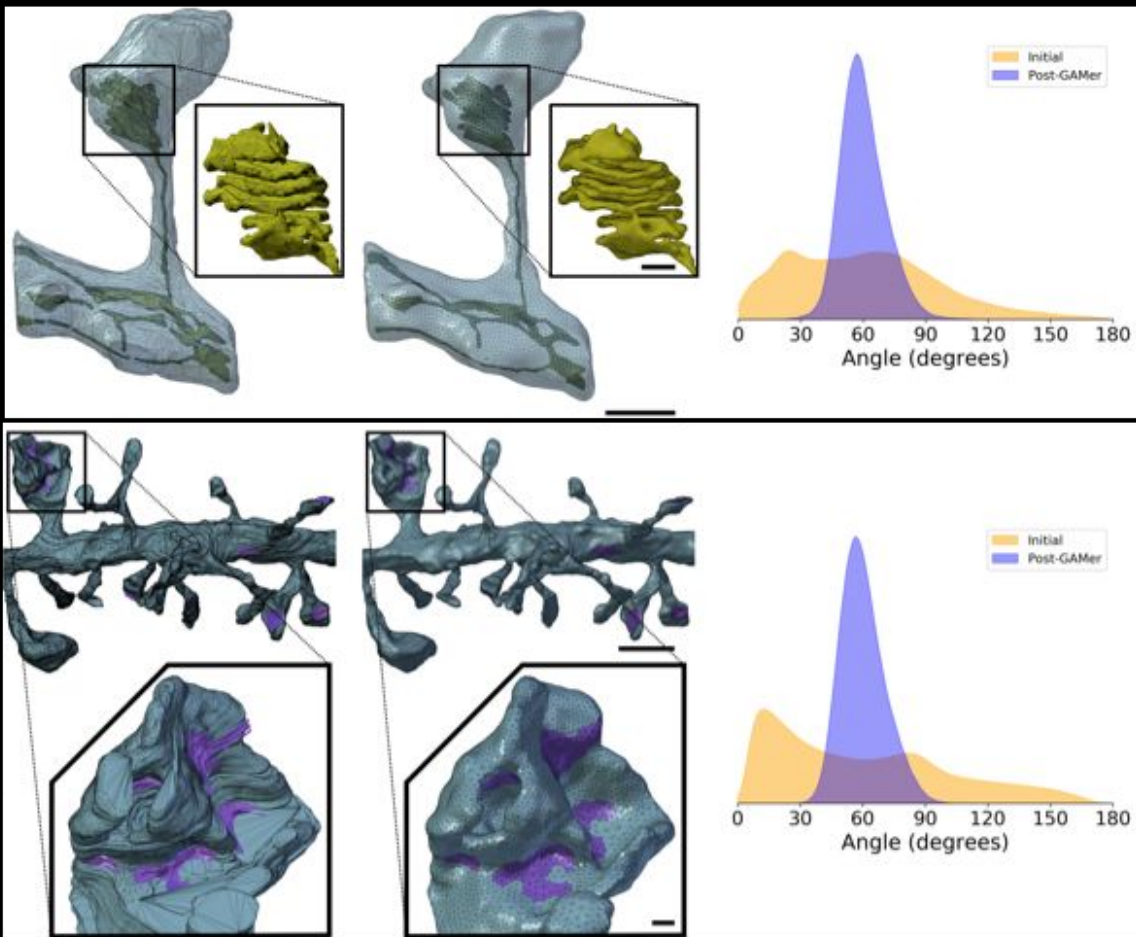


GAMer 2: Geometry-Preserving Adaptive MeshER

 [ctlee/gamer](https://github.com/ctlee/gamer)

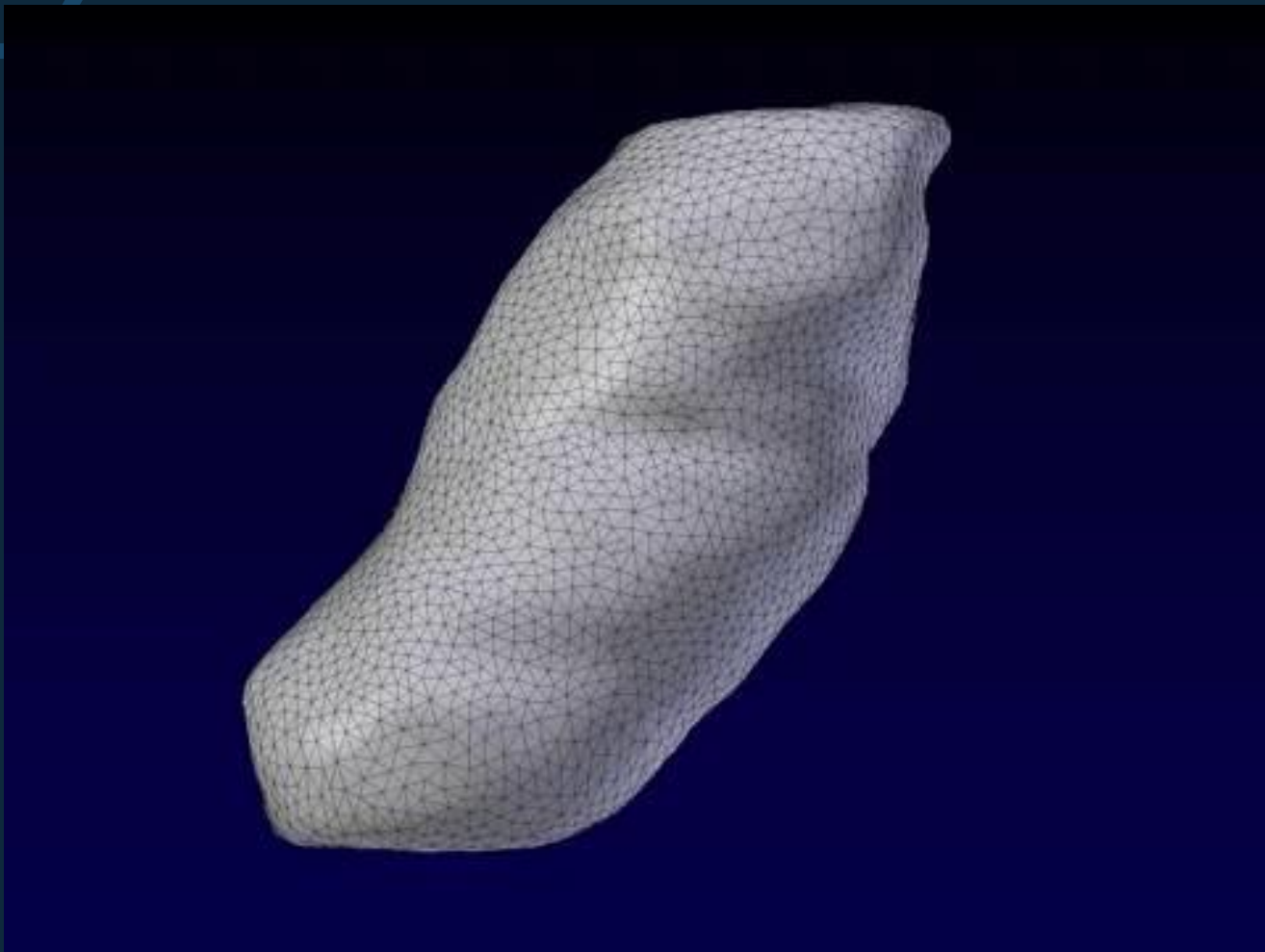
LGPL v 2.1 C++ Code

- Mesh conditioning
- Boundary marking
- Tetrahedralization (TetGen)
- Built off CASC data structure

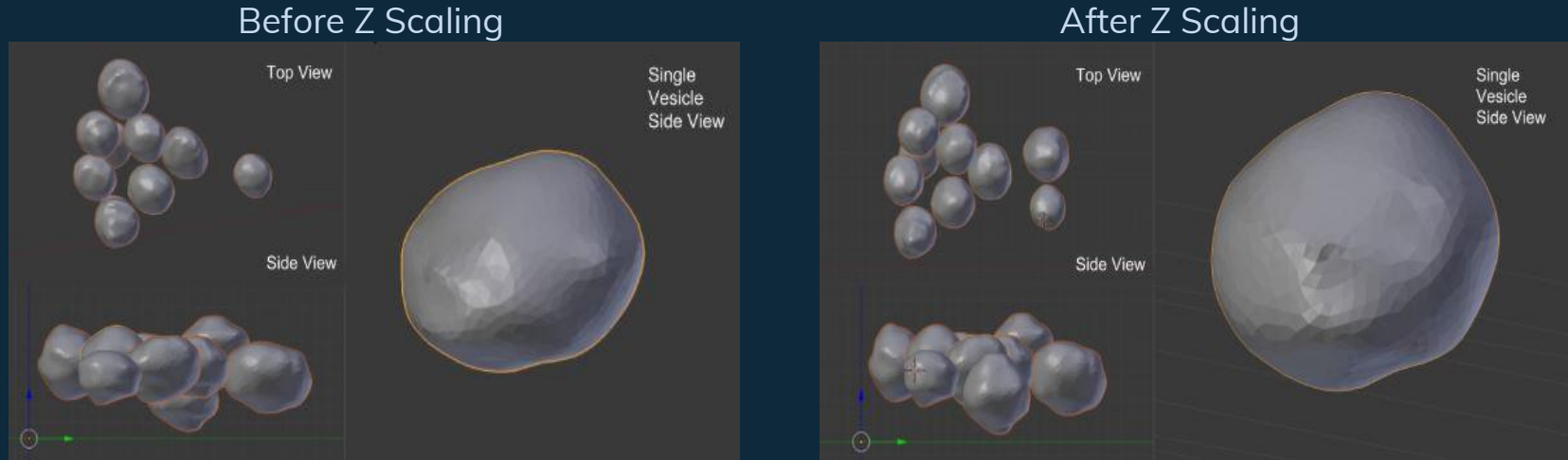


CTL*; Laughlin, JG*; et al. BioRxiv 2019.

CASC: CTL*; Moody, JB*; ACM TOMS 2019, In Press



Correcting for shrinkage using sphericity of vesicles

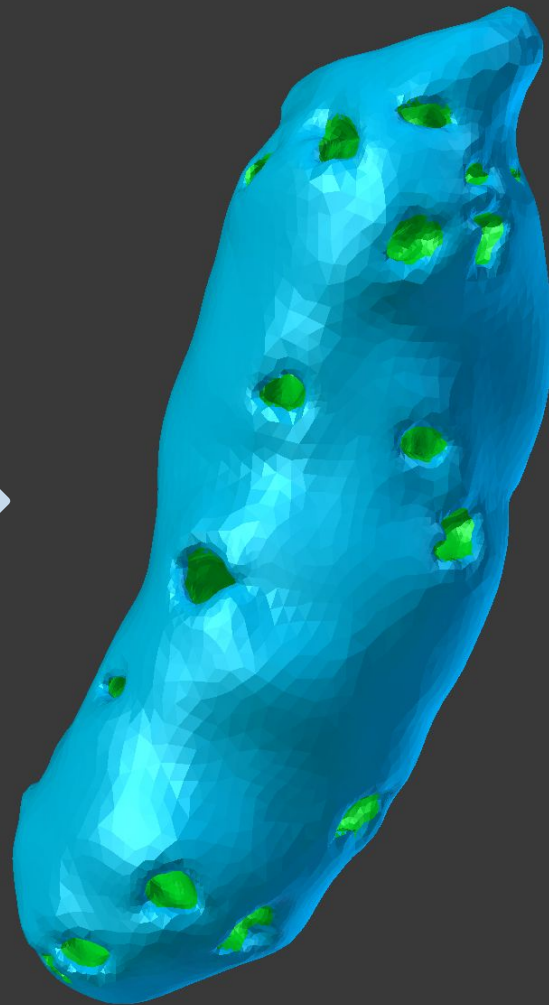
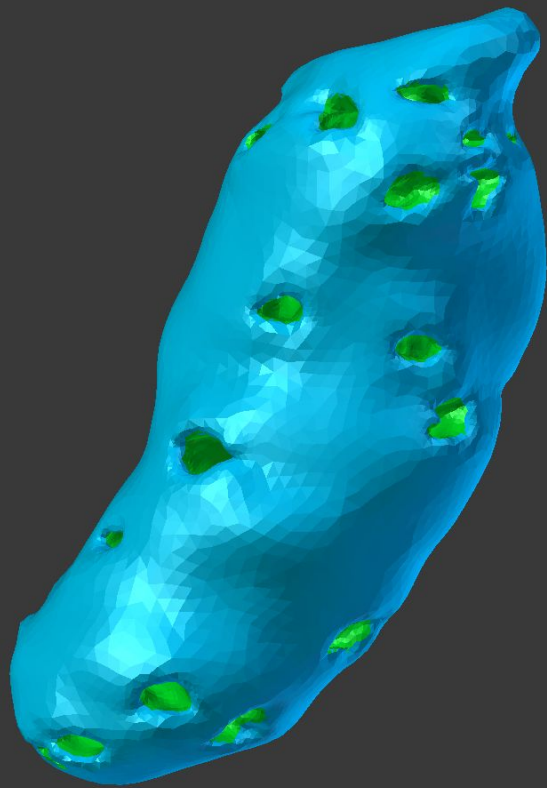


Scaling by 120% restores spherical vesicles.

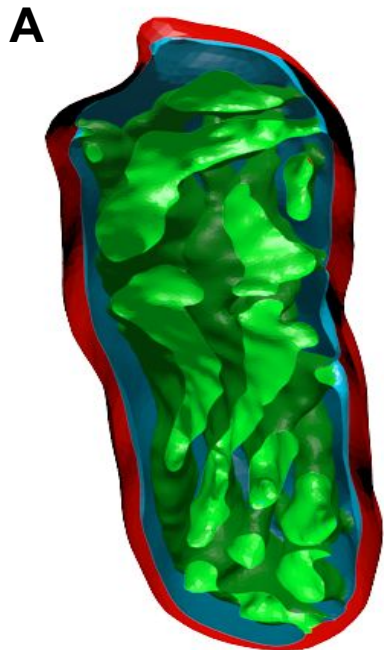
| Diameters in nm | Mito 1 | Mito 2 | Mito 3 |
|------------------|--------|--------|--------|
| X | 47 | 47 | 48 |
| Y | 51 | 50 | 53 |
| Z Before Scaling | 40 | 42 | 39 |
| Z After Scaling | 48 | 50 | 47 |



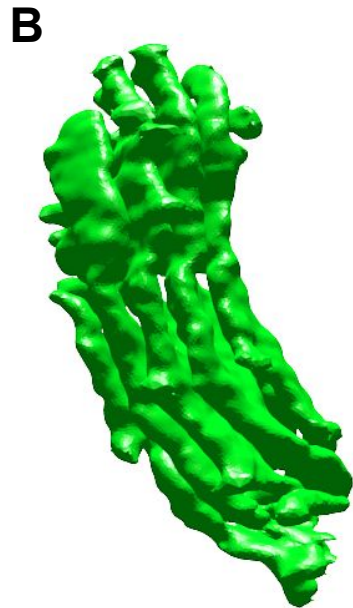
Before and After Scaling



Structure

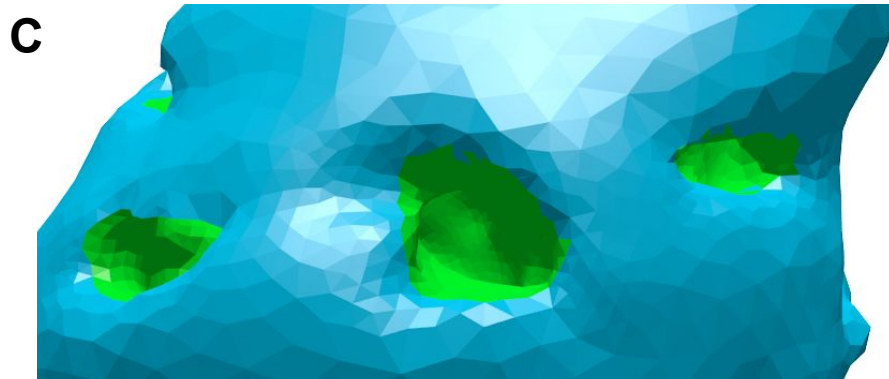


Cutaway of All Membranes

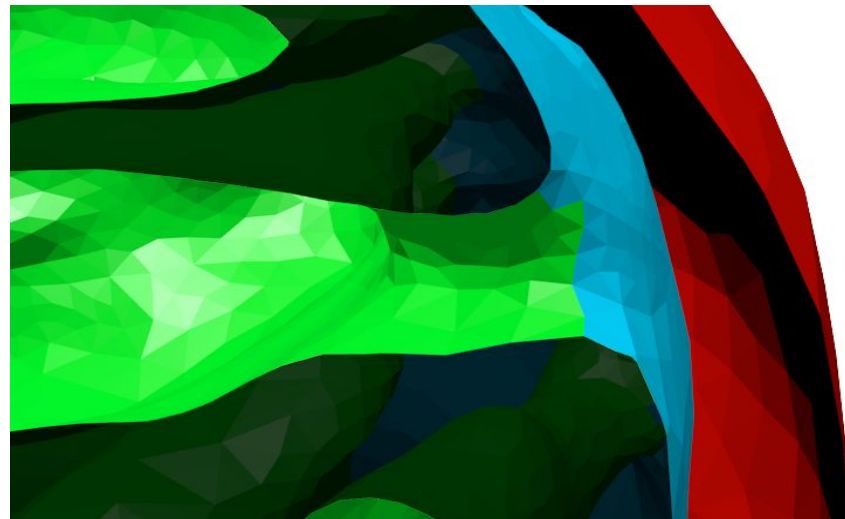


Cristae Displaying Lamellar Sheets

Cristae Junctions in the Inner Membrane



D Cutaway of a Cristae Junction



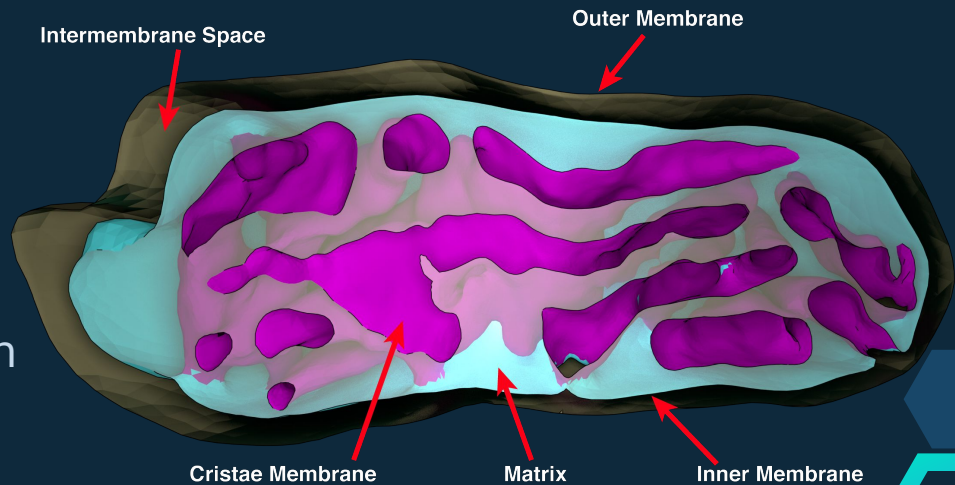
Serial Transmission
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Analysis +
Modeling

- ◇ Compartment Volumes
 - Matrix, Intercristal spaces
- ◇ Surface Area
- ◇ Membrane curvature estimation
 - Mean, Gaussian, k_1 , k_2





Cristae Connectivity

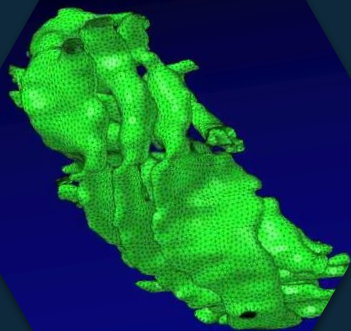
Connectivity for Adaptability

- ◇ There are no intracristal spaces which close before connecting to the rest of the network,
- ◇ There are no intracristal compartments without crista junctions.
- ◇ Free diffusion throughout the cristae compartment allows for efficient response to metabolic conditions.

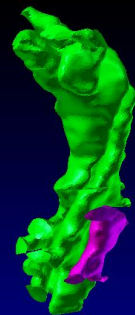
Fragmentation as a Sign of Reorganization

- ◇ Fragmentation is likely coupled to fission and fusion events.

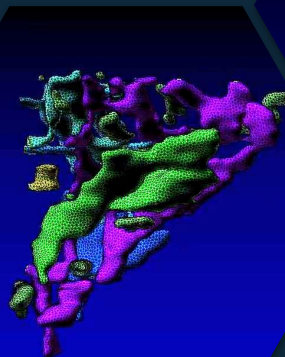
Mito 2



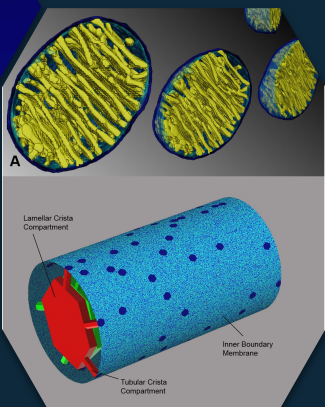
Mito 4



Mito 3



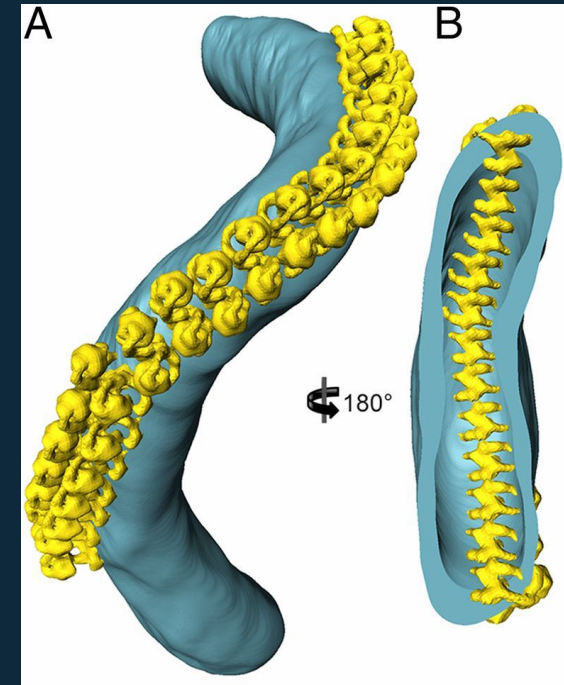
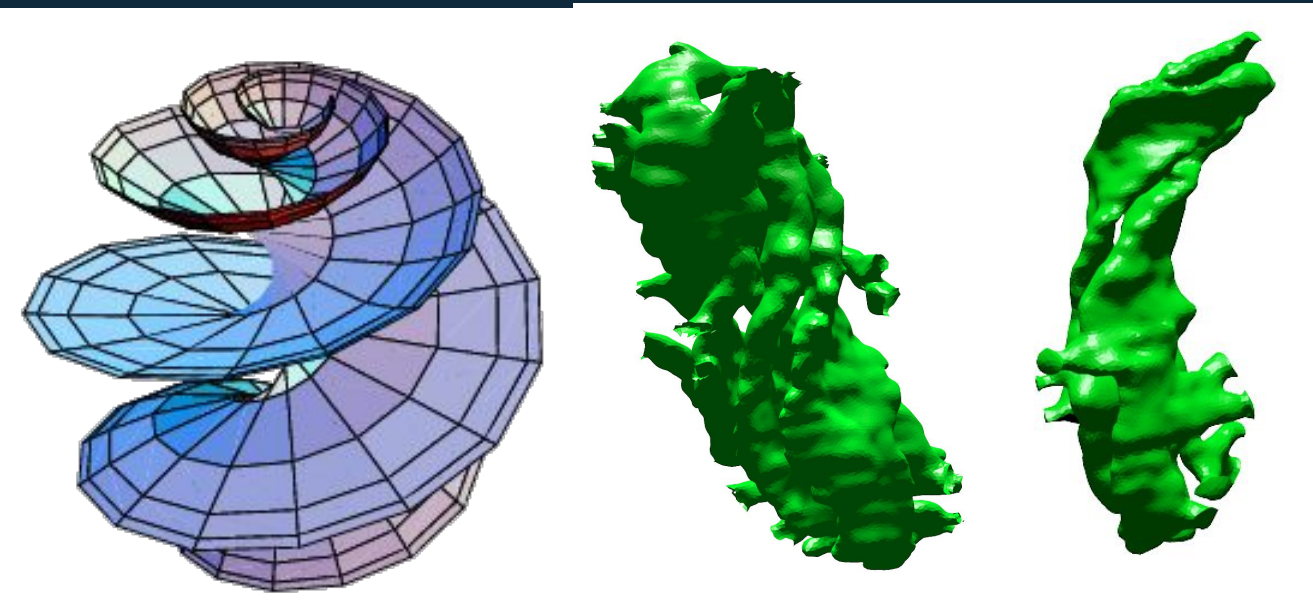
(Sukhorukov and Bereiter-Hahn, 2009)



Membrane Motifs: Helicoidal Twist

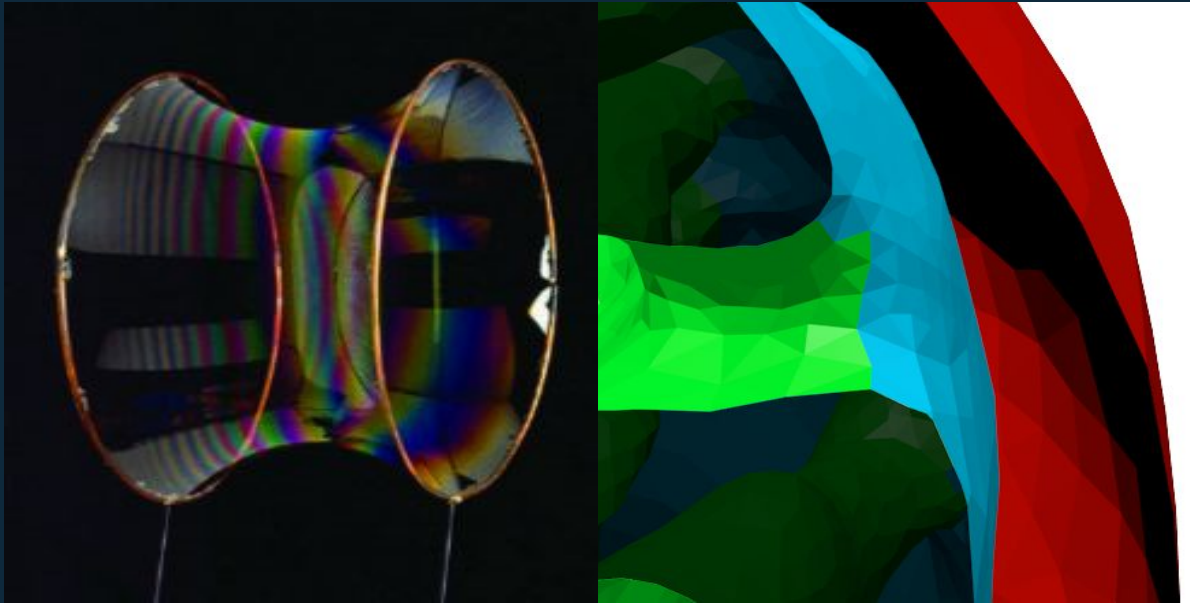
- ◇ Maximization of Space with Little Energy Expenditure
- ◇ Same Minimal Surface Motif Found in ER Sheets (Terasaki et al, 2017)
- ◇ All 4 Currently Meshed Mitochondria Exhibit Right-Handed Curves

ATP-Synthase suggested to be responsible
for RH helix in ciliate mitochondria
Muhleip, AW; et al; PNAS 2016

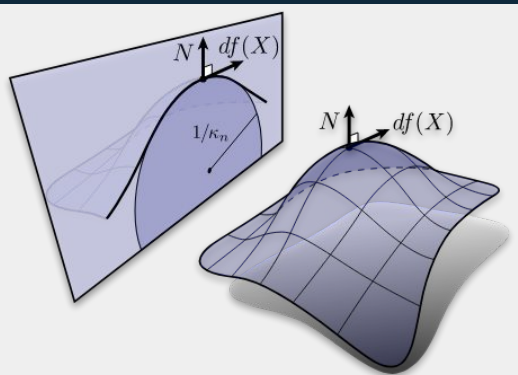


Membrane Motifs: Catenoid Cristae Junctions

- ◇ Catenoids are minimal surfaces also seen in endo- and exocytosis throughout the cell. (Chabanon and Rangamani, 2018)
- ◇ Present at cristae junctions.



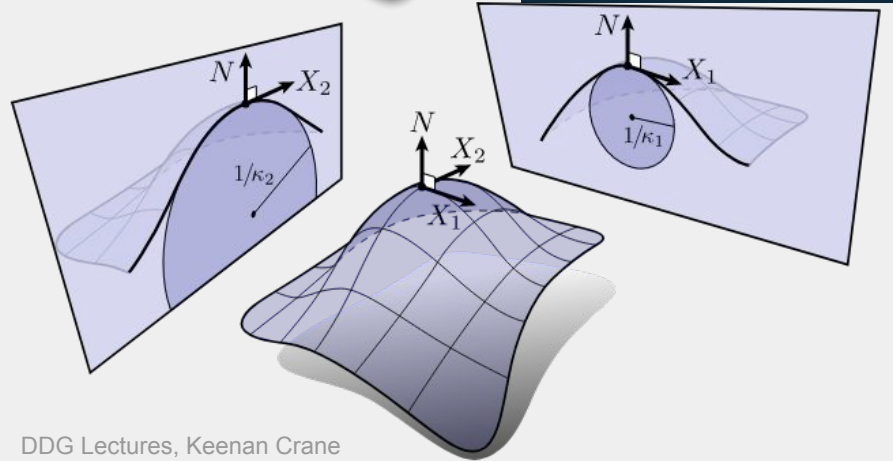
Curvatures estimated using discrete differential geometry on the smooth mesh



$$\mathbf{K}(\mathbf{x}_i) = \frac{1}{2\mathcal{A}_{\text{Mixed}}} \sum_{j \in N_1(i)} (\cot \alpha_{ij} + \cot \beta_{ij})(\mathbf{x}_i - \mathbf{x}_j)$$

Mean Curvature

$$\kappa_H = \frac{1}{2} |\mathbf{K}(\mathbf{x}_i)| = \frac{1}{2} (\kappa_1 + \kappa_2)$$



Gaussian Curvature

$$\kappa_G(\mathbf{x}_i) = (2\pi - \sum_{j=1}^{\#f} \theta_j) / \mathcal{A}_{\text{Mixed}}$$

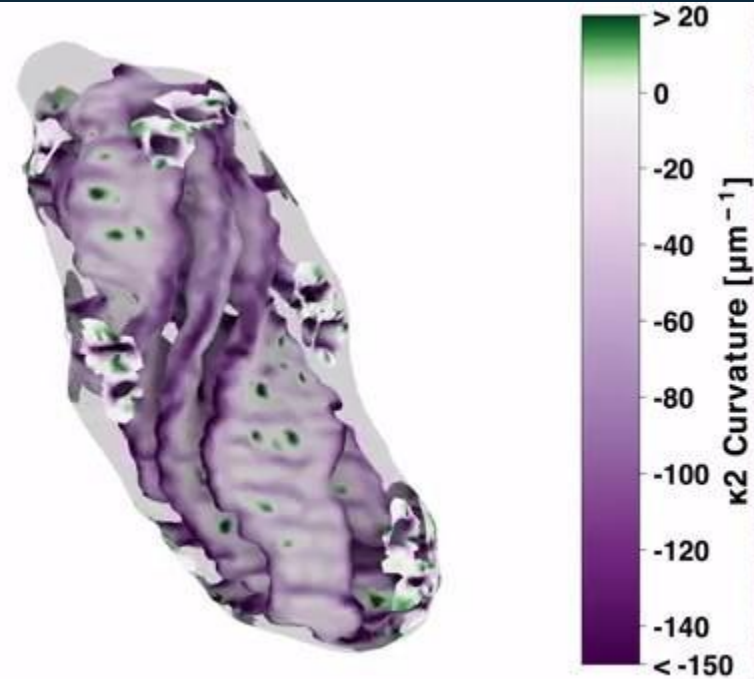
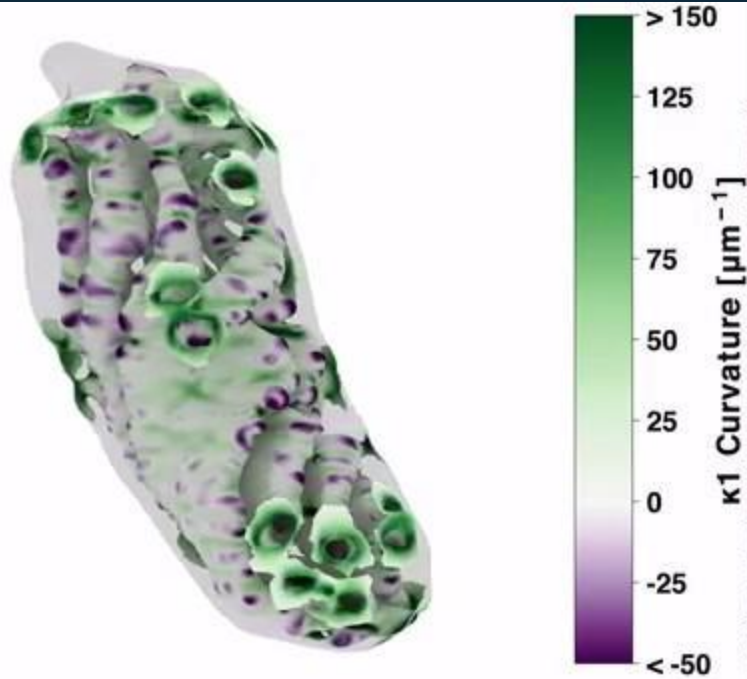
$$= \kappa_1 \kappa_2$$

First and Second Principal Curvatures

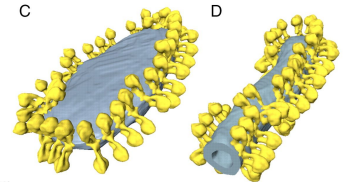
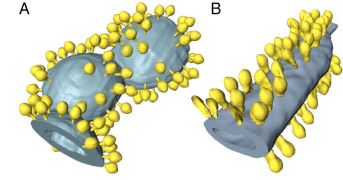
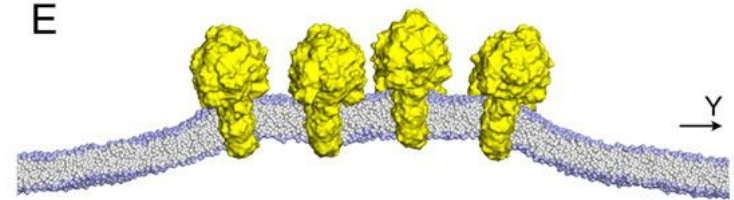
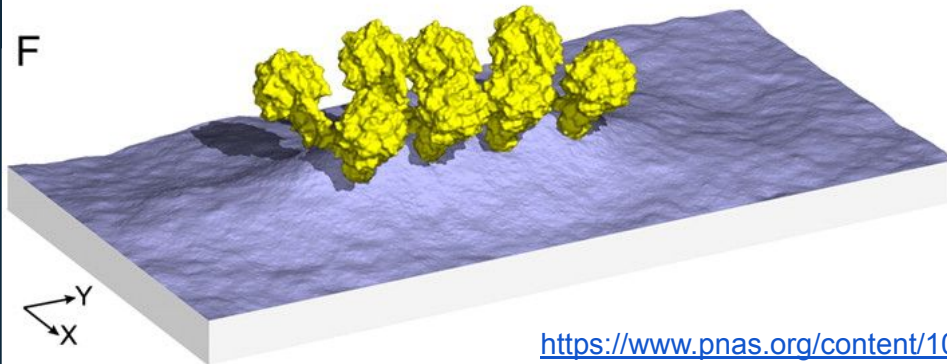
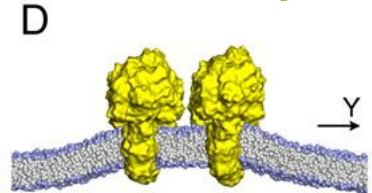
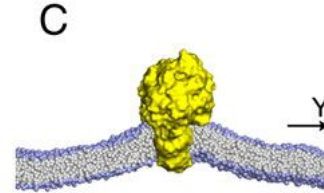
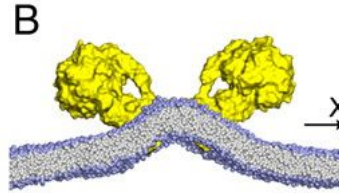
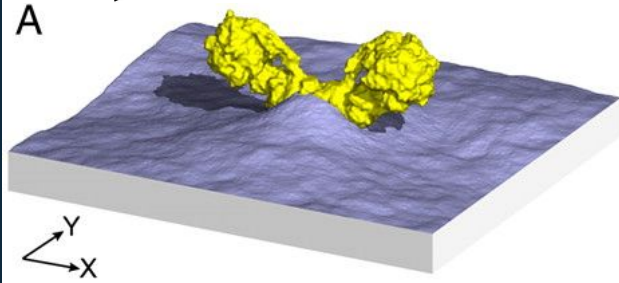
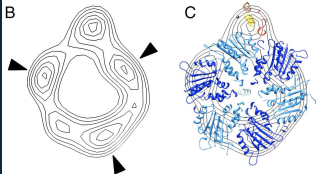
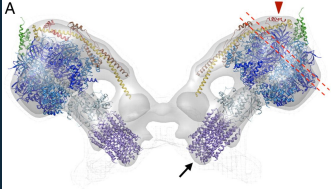
$$\kappa_1(\mathbf{x}_i) = \kappa_H(\mathbf{x}_i) + \sqrt{\Delta(\mathbf{x}_i)}$$

$$\kappa_2(\mathbf{x}_i) = \kappa_H(\mathbf{x}_i) - \sqrt{\Delta(\mathbf{x}_i)}$$

Curvatures estimated using discrete differential geometry on the smooth mesh



Cristae curvatures are often induced by ATP Synthase



<https://www.pnas.org/content/109/34/13602.short>

Dimers of mitochondrial ATP synthase induce membrane curvature and self-assemble into rows

January 25, 2018

SHORT REPORTS

Mitochondria are physiologically maintained at close to 50 °C

Dominique Chrétien^{1,2}, Paule Bénit^{1,2}, Hyung-Ho Ha³, Susanne Keipert⁴, Riyad El-Khoury⁵, Young-Tae Chang⁶, Martin Jastroch⁴, Howard T. Jacobs^{7,8}, Pierre Rustin^{1,2,9*}, Malgorzata Rak^{1,2,9}

January 25, 2018

PRIMER


Hot mitochondria?

Nick Lane*

A critique of methods for temperature imaging in single cells

Guillaume Baffou, Hervé Rigneault, Didier Marguet & Ludovic Jullien

Using 3D Models to Study Thermal Properties of Mitochondria



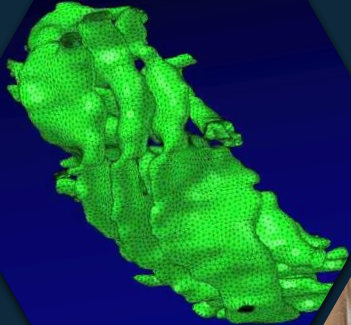
Thermodynamics & Cristae

Prevalence of Negative Gaussian Curvature

- ◇ Saddle points and areas of negative gaussian curvature facilitate large thermal fluctuations. (Evans et al., 2017)

High Surface Area to Volume Ratio

- ◇ Cristae channels maximize surface area to catalyse reactions.
- ◇ Maximum surface area leads to maximum heat distribution from the mitochondria outwards.




$$\frac{\partial \phi(\mathbf{r}, t)}{\partial t} = D \nabla^2 \phi(\mathbf{r}, t),$$



Summary

- ◇ 3D Mitochondria Models Useful for Morphological Analysis + Simulations
- ◇ Workflow produces meshes amenable to basic morphological analysis

Moving Forward

- ◇ Improving workflow throughput
 - ◇ Priya + Meagan working on ML approaches to segmentation
 - ◇ Optimizing curvature calculations
 - ◇ Creating a Mechanical Model of Mitochondrial Curvature
- 

Terry Sejnowski (Salk)

Padmini Rangamani (UCSD)

Electron Tomography

Mark Ellisman (UCSD)

Alexander Skupin (LCSB)

Guadalupe Garcia (LCSB)

Guy Perkins

Eric Bushong

Sebastien Phan

General Discussion

Justin Laughlin (UCSD)

Rangamani Lab

Sejnowski Lab

Harris Lab (UT Austin)

Segmentation and Meshing

Tom Bartol (Salk)

Don Spencer (Salk)

Priya Khandelwal (Salk)

Emily Liu (Salk)

Kelly Brockmeyer (Salk)

Aranza Sofia Martinez Lopez (UCSD,
ENLACE)

Andrea Santiago Jacinto (UCSD,
ENLACE)

Justin Oshiro (UCSD)

Meagan Rowan (UCSD)

Andrew Nguyen (UCSD)

Bob Kuczewski (Salk)

salk
Where cures begin

Meshathon Participants



UC San Diego

JACOBS SCHOOL OF ENGINEERING

**AFOSR, NSF DMS, NIH NIGMS,
Hartwell Foundation**

Thank You!

Salk Institute

Terry Sejnowski

Tom Bartol

Don Spencer

Bob Kuczewski

Priya Khandelwal

Emily Liu

Kelly Brockmeyer

UT Austin

Kristen Harris

Harris Lab

University of Luxembourg

Alexander Skupin

Guadalupe Garcia

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Eric Bushong

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Guy Perkins

Aranza Sofia Martinez Lopez

Andrea Santiago Jacinto

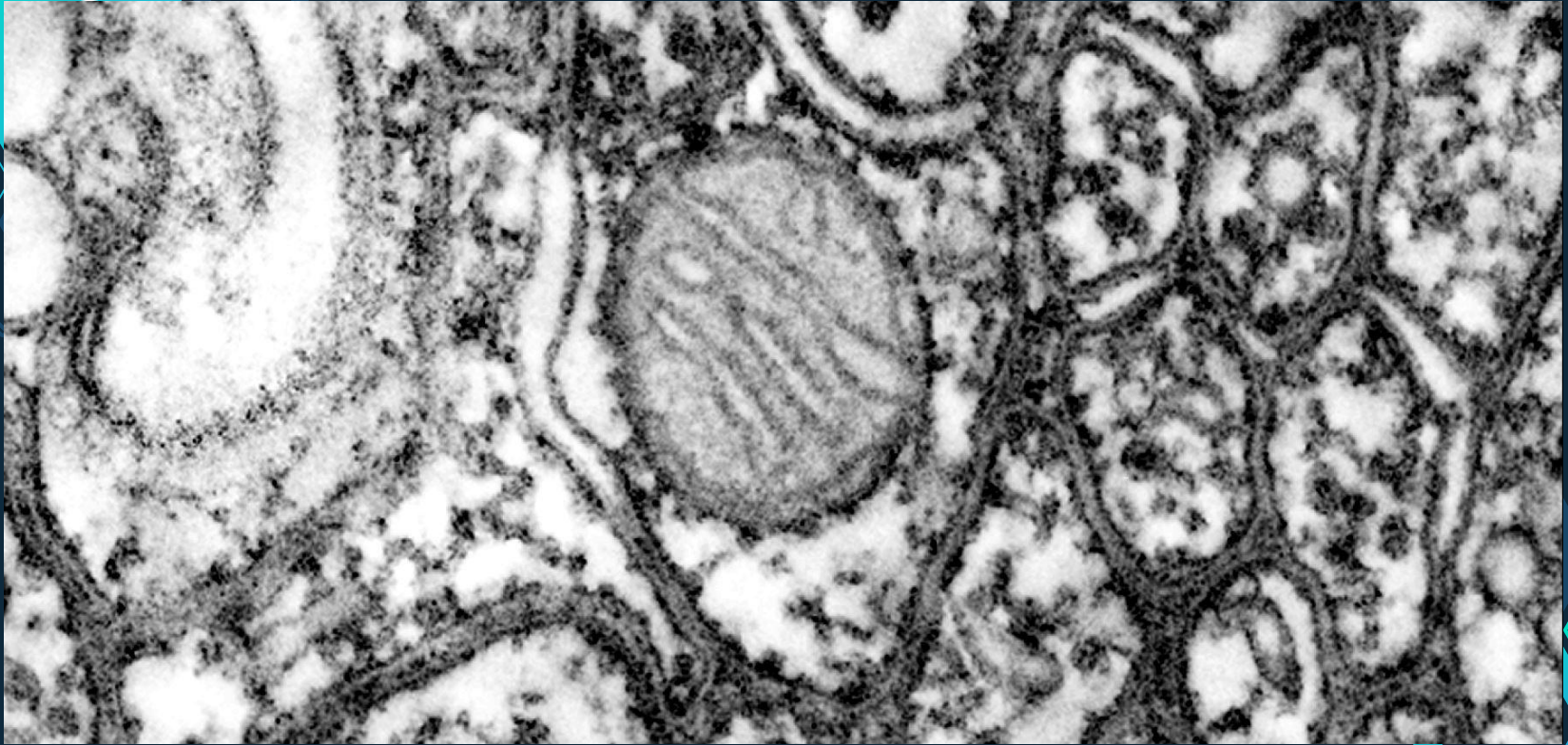
Justin Oshiro

Meagan Rowan

Andrew Nguyen

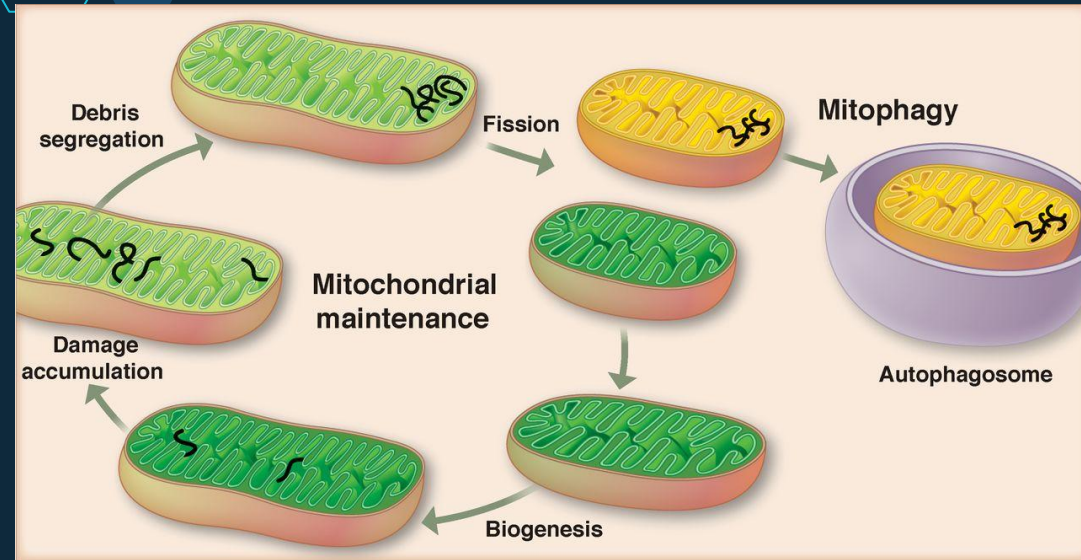
Rangamani and Ellisman Labs

**AFOSR, NSF DMS, NIH NIGMS,
Hartwell Foundation**



A Slice from a Mouse Cerebellum

A Mitochondrial Network



(Youle, 2012)

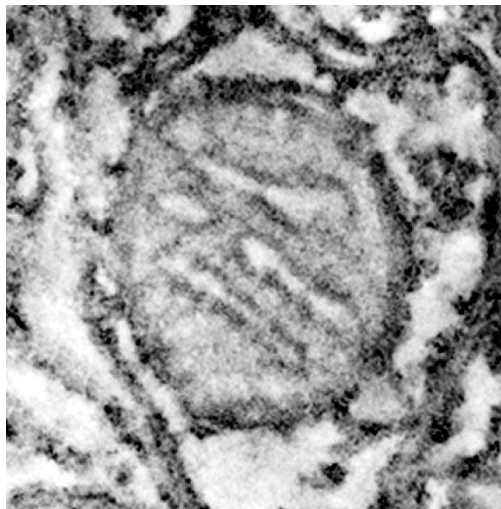
- ◇ Mitochondria can Combine (Fusion) and Divide (Fission)
- ◇ Fusion
 - Strengthens ATP Production
 - Reduces Impact of mtDNA Mutation
- ◇ Fission
 - Creates of New Mitochondria
 - “Pushes Out” Damage and Debris



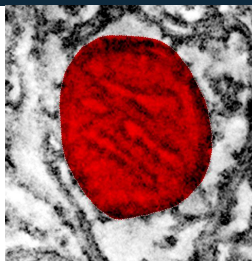
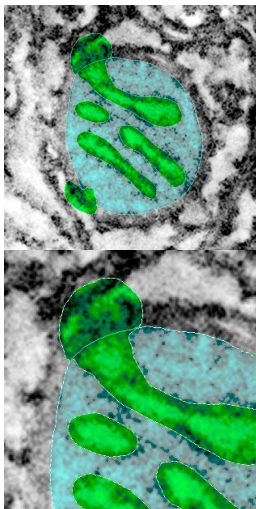
Creating a 3D Reconstruction

Workflow

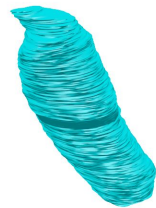
A) Tomography Images



B) Traces



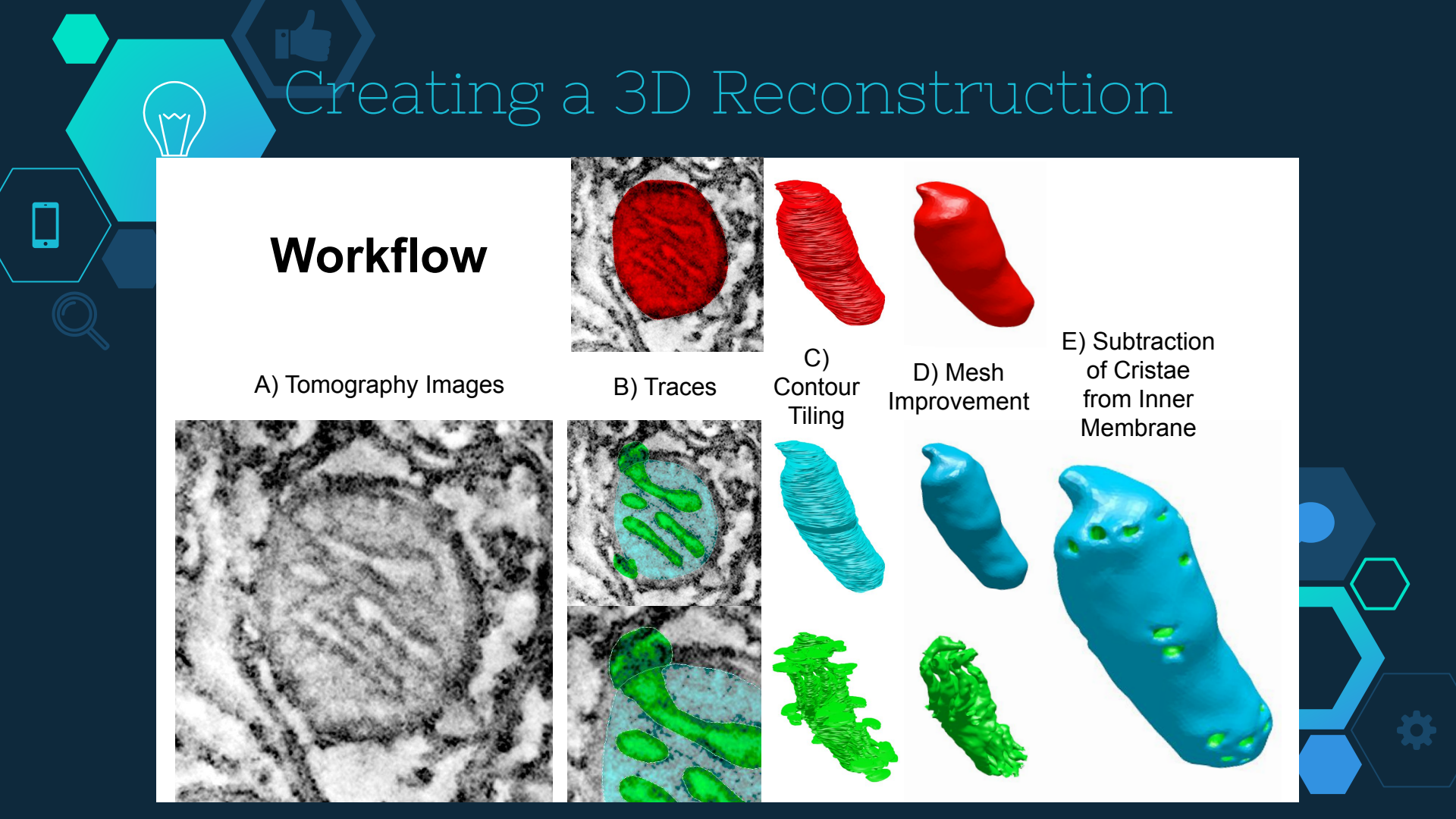
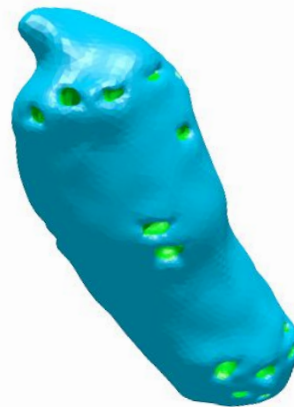
C) Contour Tiling

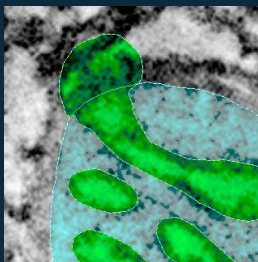
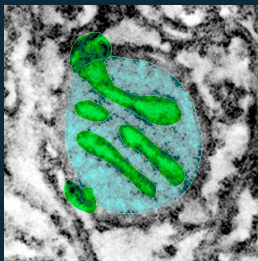
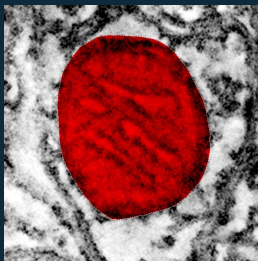
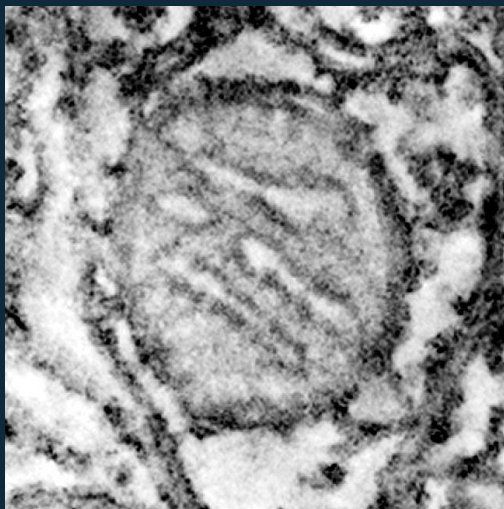


D) Mesh Improvement



E) Subtraction of Cristae from Inner Membrane



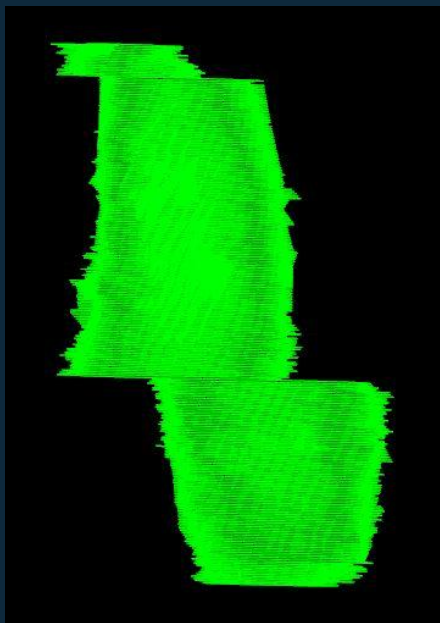


Serial Transmission
EM Tomograms of
Mouse Cerebellum

Image
Segmentation

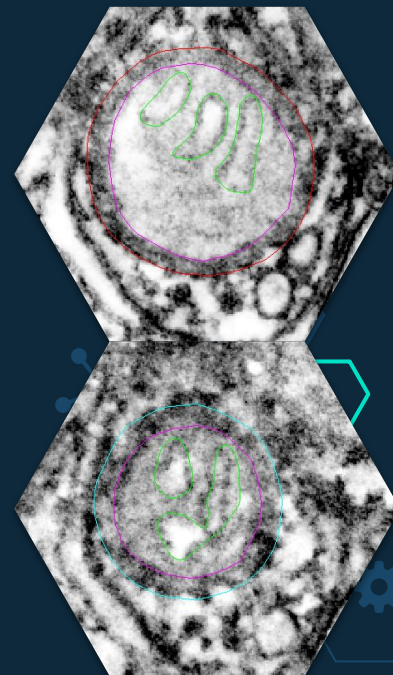
Convert to a
3D Model
with
CellBlender
and GAMer

Analysis +
Modeling



Gap Interpolation

- ◇ Physical slices of a rat brain are cut 250 nanometers thick. → CT Scans create 1.6-nanometer thick virtual slices
- ◇ Knife Removes Material Between Tomograms
- ◇ Thickness Disparity → Gaps in the Data Set
- ◇ 1.6 nm Slices, 16-20 nm Gaps
- ◇ Must be Manually Interpolated with the Help of IMOD's Interpolator





GAMer

- ◇ Designed by Zeyun Zu with the Holst Research Group
- ◇ Translates Reconstruct Traces into a Smooth Surface Model

CellBlender

- ◇ Designed by Tom Bartol for Cell Modeling
- ◇ Subtracts Cristae from Inner Membrane to form Invaginations

